



# Enantioselective synthesis of (*S*)-4-methyleneglutamic acid via tandem conjugate addition–elimination under phase-transfer catalytic conditions

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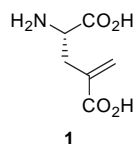
## ABSTRACT

An efficient enantioselective synthetic method for (*S*)-4-methyleneglutamic acid is reported. Phase-transfer catalytic conjugate addition–elimination of the benzophenone imine of glycine *tert*-butyl ester in the presence of chiral *Cinchona*-derived catalysts give the corresponding conjugated addition products (>99% ee), which could be hydrolyzed to chiral (*S*)-4-methyleneglutamic acid.

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## 1. Introduction

(*S*)-4-Methyleneglutamic acid (**1**), first isolated from germinated peanuts in 1951 and also found in a variety of other plants,<sup>1</sup> has been known to possess inhibitory activity against the central nervous system.<sup>2</sup> The peptide composed of Boc-4-methyleneGlu-Glu-Val has been shown to be a strong inhibitor of vitamin K-dependent carboxylation, an important process in blood clotting.<sup>3</sup>



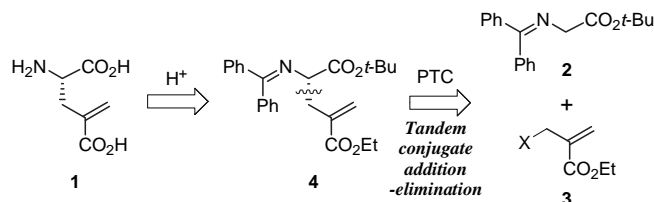
(*S*)-4-Methylene glutamic acid

As a part of our research program to study the structure–activity relationship of **1** and its derivatives, an efficient and practical synthetic method of **1** is need to be developed. Although there have been several synthetic methods for optically enriched **1**, all of the reported processes employed chiral substrates or enzymes for resolution, which might not be suitable for large scale preparation.<sup>4</sup> In the last decade, we have made efforts toward the development of novel cinchona alkaloid-derived chiral phase-transfer catalysts and their application in the enantioselective preparation

of natural and nonnatural optically active amino acid derivatives, including the asymmetric alkylation of the *N*-(diphenylmethylene) glycine *tert*-butyl ester (**2**), a powerful route toward chiral amino acids.<sup>5</sup> In this article, an efficient catalytic enantioselective synthesis of **1** via asymmetric tandem conjugate addition–elimination under phase-transfer conditions is reported.

## 2. Results and discussion

In 2005, Ramachandran et al. reported quite excellent enantioselective synthetic method for 4-alkylidenyl glutamic acid methyl esters by tandem conjugate addition–elimination of activated allylic acetates under chiral PTC conditions.<sup>6</sup> Similarity of structures led us to adapt their method for the synthesis of (*S*)-4-methyleneglutamic acid. We envisioned that an asymmetric phase-transfer catalytic alkylation can be employed as a key step, as shown in the retrosynthetic analysis (Scheme 1), for introduction of the *S* chirality, followed by acidic hydrolysis to afford **1**.

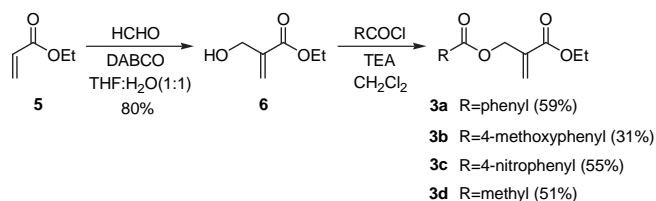


Scheme 1. Strategy for the synthesis of (*S*)-4-methyleneglutamic acid (**1**).

Michael acceptors (**3a–d**) were prepared in two steps from ethyl acrylate (**5**) by the Baylis–Hillman reaction,<sup>7</sup> followed by

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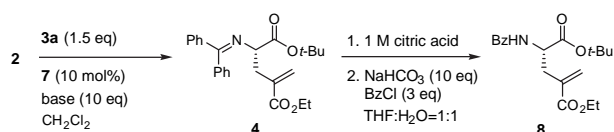
O-acylation (Scheme 2). Treatment of ethyl acrylate (**5**) with formaldehyde in the presence of DABCO in THF/H<sub>2</sub>O (1:1) afforded ethyl 2-(hydroxymethyl) acrylate (**6**) in 80% yield. Acylation of **6** with the selected acid chlorides gave the corresponding Michael acceptors (**3a–d**).



Scheme 2. Preparation of Michael acceptors.

Catalytic asymmetric conjugate addition–elimination was carried out with ethyl 2-(acyloxymethyl)acrylate **3** and benzophenone imine of glycine ester **2** under phase-transfer conditions (Table 1). Four kinds of hydrocinchonidinium salts (**7a–d**, Fig. 1),<sup>8</sup> which have previously shown excellent capabilities as chiral phase-transfer catalysts in the asymmetric alkylation of **2**, were employed for chirality generation.

Table 1  
Enantioselective tandem conjugate addition–elimination under PTC conditions<sup>a</sup>



Entry	<b>7</b>	Base	Temp (°C)	Time <sup>b</sup> (h)	Yield <sup>c</sup> (%)	ee (%) <sup>d</sup> (config.) <sup>e</sup>
1	<b>7a</b>	50% KOH	–40	16	93	64 (S)
2	<b>7b</b>	50% KOH	–40	20	87	78 (S)
3	<b>7c</b>	50% KOH	–40	16	86	76 (S)
4	<b>7d</b>	50% KOH	–40	26	84	90 (S)
5	<b>7d</b>	50% KOH	–55	30	85	92 (S)
6	<b>7d</b>	CsOH·H <sub>2</sub> O	–78	12	91	92 (S)

<sup>a</sup> Reaction conducted with the benzophenone imine of glycine *tert*-butyl ester (**2**, 1.0 equiv), Michael acceptor (**3a**, 1.5 equiv), base (10.0 equiv), and PTC (**7**, 0.1 equiv).

<sup>b</sup> Reaction time of the first step.

<sup>c</sup> Isolated yield for the two steps.

<sup>d</sup> Enantiopurity determined by chiral HPLC analysis of *N*-benzoyl derivative **8** using a chiral column (DAICEL Chiralpak AD-H) with hexane/2-propanol as the solvent system.

<sup>e</sup> Absolute configuration determined by comparing with the optical rotation value of the 4-methyleneglutamic acid HCl salt, obtained by hydrolysis of **8** in 6.0 M HCl, with literature value.<sup>4c</sup>

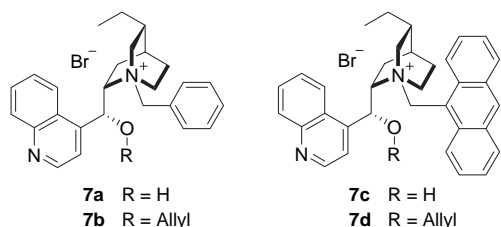


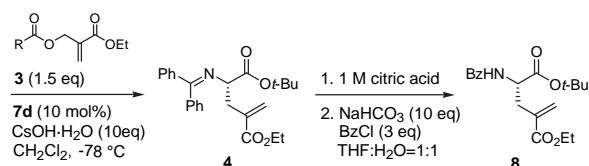
Figure 1. Phase-transfer catalysts.

Screening of PTC was first performed under the same conditions (3a Michael acceptor, 50% KOH base, CH<sub>2</sub>Cl<sub>2</sub> solvent, and –40 °C reaction temperature), and the results are shown in Table 1. The *O*-allyl-*N*-(9-anthracenylmethyl) hydrocinchonidinium bromide (**7d**) gave the product with the highest enantioselectivity (entry 4, 84% yield, 90% ee). Generally, C(9)-*O*-allyl catalysts (entry 2, **7b**, 78%

ee; entry 4, **7d**, 90% ee) showed higher enantioselectivities compared to those of C(9)-*OH* catalysts (entry 1, **7a**, 64% ee; entry 3, **7c**, 76% ee), respectively. Further tuning of the reaction conditions revealed that use of the CsOH·H<sub>2</sub>O base at lower reaction temperature (–78 °C) was highly advantageous to improve both chemical and optical yields (entry 6, 91% yield, 92% ee).

Next, the Michael acceptor was optimized under the best reaction conditions (entry 6 in Table 1) to improve the enantioselectivity, which was not studied in the works of Ramachandran et al.<sup>6</sup> Asymmetric tandem conjugate addition–elimination of **2** with the prepared Michael acceptors (**3b–d**) were performed under reaction conditions of CsOH·H<sub>2</sub>O in the presence of **7d** in dichloromethane at –78 °C (Table 2). Chemical yield and enantioselectivity were variable depending upon the leaving groups in the Michael acceptors. All afforded the conjugate addition–elimination product (**4**) in high enantioselectivities. It was expected that **3c**, with a better leaving group, would provide a higher chemical yield, but the lowest yield with moderate enantioselectivity was observed among the Michael acceptors (**3a–d**). The 4-methoxy-incorporated **3b** showed chemical and optical yields substantially higher than 4-nitro analogue **3c**. The smaller acetoxy group (**3d**) gave the highest chemical yield. The highest enantioselectivity was obtained with **3b** (entry 2, 99% ee). There is no clear evidence yet but we speculate that the 4-methoxybenzoyl group might play an important role for the high enantioselectivity in electronically or spatially in PTC reaction process. Hydrolysis of **8** (99% ee) in 6.0 M HCl gave the (*S*)-4-methyleneglutamic acid hydrochloride salt (**1**·HCl) in 97% yield. Absolute configuration was confirmed as *S* by comparing optical rotation with the reported value [ $[\alpha]_D^{20} +13.5$  (c 0.5, HCl); lit.<sup>4c</sup> [ $[\alpha]_D^{20} +13.2$  (c 1.0, HCl)]}.

Table 2  
Enantioselective phase-transfer catalytic Michael reaction and elimination reaction<sup>a</sup>



Entry	<b>3</b>	R	Time <sup>b</sup> (h)	Yield <sup>c</sup> (%)	ee (%) <sup>d</sup> (config.)
1	<b>3a</b>	Phenyl	12	91	92 (S)
2	<b>3b</b>	4-Methoxyphenyl	14	90	99 (S)
3	<b>3c</b>	4-Nitrophenyl	13	77	93 (S)
4	<b>3d</b>	Methyl	12	95	96 (S)

<sup>a</sup> Reactions carried out with benzophenone imine of glycine *tert*-butyl ester **2** (1.0 equiv), Michael acceptor **3** (1.5 equiv), CsOH·H<sub>2</sub>O (10.0 equiv), **7d** (0.1 equiv).

<sup>b</sup> Reaction time of the first step.

<sup>c</sup> Isolated yield for the two steps.

<sup>d</sup> Enantiopurity and absolute configuration were determined as Table 1.

### 3. Conclusions

A new enantioselective synthetic method of (*S*)-4-methyleneglutamic acid (**1**) under asymmetric phase-transfer catalytic tandem conjugate addition–elimination of the benzophenone imine of glycine *tert*-butyl ester (**2**) with chiral cinchona catalysts has been developed. Highly optically active **1**·HCl was prepared from **2** in three steps at 88% chemical yield and 99% ee.

### 4. Experimental

#### 4.1. General

Infrared (IR) spectra were recorded on a JASCO FT/IR-300E and Perkin–Elmer 1710 FT spectrometer. <sup>1</sup>H NMR was measured at 300

or 400 MHz and  $^{13}\text{C}$  NMR at 75, 100, or 125 MHz. Chemical shifts were recorded in parts per million relative to  $\text{CHCl}_3$  ( $\delta$  7.24) and  $\text{H}_2\text{O}$  ( $\delta$  4.70) for  $^1\text{H}$  NMR and relative to the central  $\text{CDCl}_3$  ( $\delta$  77.21) resonance for  $^{13}\text{C}$  NMR. Melting points were not corrected. Enantiomeric excess (ee) was determined by HPLC using 4.6 mm $\times$ 250 mm Daicel Chiralpak AD-H column. Optical rotations were measured on a JASCO DIP-1000 digital polarimeter. High-resolution mass spectra (HRMS) were measured on a JEOL JMS-AX 505wA, JEOL JMS-HX/HX 110A spectrometer. Flash column chromatography was carried out using silica gel (230–400 mesh). For thin-layer chromatography (TLC) analysis, Merck precoated TLC plates (silica gel 60 GF<sub>254</sub>, 0.25 mm) were used. All solvents and commercially available chemicals were used without additional purification. Ethyl 2-(hydroxymethyl)acrylate (**6**) was prepared according to reported procedures.<sup>7</sup>

#### 4.2. Ethyl 2-(benzoyloxymethyl)acrylate (**3a**)

To a cooled solution of ethyl 2-(hydroxymethyl)acrylate (**6**) (1.0 g, 7.68 mmol) in dichloromethane (30 mL) were added triethylamine (1.61 mL, 11.52 mmol) and benzoyl chloride (1.34 mL, 11.52 mmol). After stirring for 1 h, the reaction mixture was washed with brine (2 $\times$ 30 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=10:1) to afford **3a** (1.06 g, 59%) as a colorless oil.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.06–8.03 (m, 2H), 7.58–7.53 (m, 1H), 7.46–7.40 (m, 2H), 6.40 (s, 1H), 5.91 (s, 1H), 5.05 (s, 2H), 4.24 (q,  $J$ =7.14 Hz, 2H), 1.29 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 165.1, 135.5, 133.0, 130.4, 129.5, 128.3, 127.0, 62.7, 14.0 ppm; IR (KBr)  $\nu$  2983, 1789, 1725, 1602, 1452, 1416, 1315, 1274, 1213, 1175, 1111, 1070, 1027, 712  $\text{cm}^{-1}$ ; HRMS (FAB): calcd for  $[\text{C}_{13}\text{H}_{15}\text{O}_4]^+$ : 235.0970, found: 235.0964.

#### 4.3. Ethyl 2-(4-methoxybenzoyloxymethyl)acrylate (**3b**)

To a cooled solution of ethyl 2-(hydroxymethyl)acrylate (**6**) (1.0 g, 7.68 mmol) in dichloromethane (30 mL) were added triethylamine (1.61 mL, 11.52 mmol) and 4-methoxybenzoyl chloride (2.62 g, 15.36 mmol). After stirring for 3 h, the reaction mixture was washed with brine (2 $\times$ 30 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=20:1) to afford **3b** (634 mg, 31%) as a white solid. Mp 40  $^\circ\text{C}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.13–7.98 (m, 2H), 6.93–6.88 (m, 2H), 6.38 (s, 1H), 5.89 (s, 1H), 5.02 (s, 2H), 4.24 (q,  $J$ =7.14 Hz, 2H), 3.85 (s, 3H), 1.28 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  165.6, 165.2, 135.7, 131.6, 126.8, 122.2, 113.6, 62.5, 60.9, 55.4, 14.1 ppm; IR (KBr)  $\nu$  2980, 1721, 1606, 1511, 1458, 1366, 1309, 1258, 1167, 1101, 1028, 848, 769, 696  $\text{cm}^{-1}$ ; HRMS (FAB): calcd for  $[\text{C}_{14}\text{H}_{17}\text{O}_5]^+$ : 265.1076, found: 265.1083.

#### 4.4. Ethyl 2-(4-nitrobenzoyloxymethyl)acrylate (**3c**)

To a cooled solution of ethyl 2-(hydroxymethyl)acrylate (**6**) (1.0 g, 7.68 mmol) in dichloromethane (30 mL) were added triethylamine (1.64 mL, 11.78 mmol) and 4-nitrobenzoyl chloride (2.18 g, 11.78 mmol). After stirring for 10 min, the reaction mixture was washed with brine (2 $\times$ 30 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=20:1) to afford **3c** (1.21 g, 55%) as a pale yellow solid. Mp 78  $^\circ\text{C}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.29–8.18 (m, 4H), 6.44 (s, 1H), 5.93 (s, 1H), 5.08 (s, 2H), 4.25 (q,  $J$ =7.14 Hz, 2H), 1.29 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.0, 164.1, 150.6, 135.2, 135.1, 130.7, 128.2, 123.5, 63.8, 61.1, 14.1 ppm; IR (KBr)  $\nu$  3114, 1725, 1648, 1517, 1460, 1406, 1348, 1272, 1117, 1009, 963, 878, 854, 819, 784, 719  $\text{cm}^{-1}$ ; HRMS (FAB): calcd for  $[\text{C}_{13}\text{H}_{14}\text{O}_6\text{N}]^+$ : 280.0821, found: 280.0813.

#### 4.5. Ethyl 2-(acetoxymethyl)acrylate (**3d**)

To a cooled solution of ethyl 2-(hydroxymethyl)acrylate (**6**) (1.0 g, 7.68 mmol) in dichloromethane (30 mL) were added triethylamine (2.75 mL, 19.74 mmol) and acetyl chloride (1.4 mL, 19.74 mmol). After stirring for 20 min, the reaction mixture was washed with brine (2 $\times$ 30 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=20:1) to afford **3d** (1.21 g, 51%) as a colorless oil.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  6.32 (s, 1H), 5.79 (s, 1H), 4.76 (s, 2H), 4.20 (q,  $J$ =7.14 Hz, 2H), 2.06 (s, 3H), 1.26 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.2, 165.0, 135.3, 127.0, 62.3, 60.8, 20.7, 14.0 ppm; IR (KBr)  $\nu$  2984, 1748, 1725, 1643, 1370, 1305, 1231, 1178, 1050, 953, 816  $\text{cm}^{-1}$ .

#### 4.6. (S)-1-tert-Butyl 5-ethyl 2-(diphenylmethylene-amino)-4-methylenepentanedioate (**4**)

A suspension of *N*-(diphenylmethylene)glycine *tert*-butyl ester (**2**) (30 mg, 0.102 mmol), ethyl 2-(4-methoxybenzoyloxymethyl)acrylate (**3b**) (40.4 mg, 0.153 mmol), and chiral phase-transfer catalyst **7d** (6.2 mg, 0.010 mmol) in dichloromethane (0.5 mL) was cooled at  $-78$   $^\circ\text{C}$ . Solid  $\text{CsOH}\cdot\text{H}_2\text{O}$  (171 mg, 1.02 mmol) was then added to the reaction mixture. After stirring at  $-78$   $^\circ\text{C}$  for 14 h, the reaction mixture was washed with brine (2 $\times$ 2 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=20:1) to afford **4** (38 mg, 90%) as a pale yellow oil.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.59–7.57 (m, 2H), 7.39–7.26 (m, 6H), 7.11–7.08 (m, 2H), 6.17 (s, 1H), 5.60 (s, 1H), 4.18 (q,  $J$ =4.38 Hz, 1H), 4.03–3.91 (m, 2H), 3.00 (dd,  $J$ =13.53, 4.2 Hz, 1H), 2.79 (dd,  $J$ =13.35, 8.97 Hz, 1H), 1.43 (s, 9H), 1.11 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.6, 170.5, 166.4, 139.5, 136.8, 136.4, 130.1, 128.7, 128.4, 128.2, 128.0, 127.9, 81.1, 64.5, 60.4, 36.0, 28.0, 27.9, 13.9 ppm; IR (KBr)  $\nu$  2978, 1721, 1626, 1446, 1392, 1368, 1280, 1151, 1028, 950, 846, 781, 698  $\text{cm}^{-1}$ ; HRMS (FAB): calcd for  $[\text{C}_{14}\text{H}_{17}\text{O}_5]^+$ : 408.2175, found: 408.2165.

#### 4.7. (S)-1-tert-Butyl 5-ethyl 2-benzamido-4-methylenepentanedioate (**8**)

A mixture of **4** (37.4 mg, 0.092 mmol), 1.0 M citric acid (1.5 mL), and tetrahydrofuran (1.5 mL) was stirred at room temperature for 2 h. Sodium bicarbonate was added to the mixture until the pH reached 7–8, and then benzoyl chloride (0.036 mL, 0.306 mmol) was added. The mixture was stirred for 5 h. The tetrahydrofuran was then evaporated and the residue diluted with ethyl acetate (3 mL). The mixture was washed with brine (2 $\times$ 3 mL), dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated. The residue was purified by column chromatography (hexanes/EtOAc=10:1) to afford **8** (32 mg, 90%) as a colorless oil.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.80–7.77 (m, 2H), 7.50–7.38 (m, 3H), 7.14 (d,  $J$ =6.96, 1H), 6.26 (s, 1H), 5.70 (s, 1H), 4.81–4.74 (m, 1H), 4.25–4.15 (m, 2H), 2.93–2.78 (m, 2H), 1.45 (s, 9H), 1.27 (t,  $J$ =7.14 Hz, 3H) ppm;  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.5, 167.2, 166.7, 136.1, 133.8, 131.4, 128.4, 126.9, 82.2, 61.1, 53.0, 34.4, 27.8, 25.2, 13.9 ppm; IR (KBr)  $\nu$  3347, 2979, 1719, 1645, 1580, 1533, 1488, 1368, 1309, 1228, 1154, 1026, 953, 846, 714  $\text{cm}^{-1}$ ; HRMS (FAB): calcd for  $[\text{C}_{19}\text{H}_{26}\text{O}_5\text{N}]^+$ : 348.1811, found: 348.1803. Enantiopurity was determined by chiral HPLC analysis (DIACEL Chiralcel AD-H, hexanes/2-propanol=9:1), flow rate=1.0 mL/min, 23  $^\circ\text{C}$ ,  $\lambda$ =254 nm, retention time, minor 10.4 min, minor 20.7 min, 99% ee.

#### 4.8. (S)-4-Methyleneglutamic acid hydrochloride (**1**·HCl)

A mixture of **8** (109 mg, 0.314 mmol) and 6.0 M HCl (2 mL) was refluxed for 24 h. Concentration of the reaction mixture in vacuo

afforded **1**·HCl (59.3 mg, 97%) as a pale yellow solid. Mp 196 °C; <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O) δ 6.39 (s, 1H), 5.89 (s, 1H), 4.31–4.16 (m, 1H), 3.75 (d, *J*=10.5, 3H), 3.01–2.79 (m, 2H) ppm; <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 171.3, 169.6, 133.6, 132.2, 52.3, 32.5 ppm; [ $\alpha$ ]<sub>D</sub><sup>20</sup> +13.2 (c 0.5, HCl); lit.<sup>4c</sup> [ $\alpha$ ]<sub>D</sub> +13.2 (c 1.0, HCl).

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### Supplementary data

Supplementary data associated with this article can be found in the online version, at doi:10.1016/j.tet.2010.04.034.

### References and notes

- (a) Done, J.; Fowden, L. *Biochem. J.* **1952**, *51*, 451; (b) Zacharius, R. M.; Pollard, J. K.; Steward, F. C. *J. Am. Chem. Soc.* **1954**, *76*, 1961; (c) Tschiersch, B. *Phytochemistry* **1962**, *1*, 103; (d) Blake, J.; Fowden, L. *Biochem. J.* **1964**, *92*, 136; (e) Watson, R.; Fowden, L. *Phytochemistry* **1973**, *12*, 617; (f) Hatanaka, S.-I.; Katayama, H. *Phytochemistry* **1975**, *14*, 1434; (g) Evans, C. S.; Bell, E. A. *Phytochemistry* **1978**, *17*, 1127; (h) Shewry, P. R.; Fowden, L. *Phytochemistry* **1976**, *15*, 1981; (i) Meier, L. K.; Sorensen, H. *Phytochemistry* **1979**, *18*, 1173; (j) Kasai, T.; Nishitoba, T.; Shiroshita, Y.; Sakamura, S. *Agric. Biol. Chem.* **1984**, *48*, 2271.
- Powell, G. K.; Dekker, E. E. *Prep. Biochem.* **1981**, *11*, 339.
- Guibé, E.; Decotignies-LeMaréchal, P.; Le Maréchal, P.; Azerad, R. *FEBS Lett.* **1984**, *177*, 265.
- (a) Baldwin, J. E.; Adlington, R. M.; Robinson, N. G. *J. Chem. Soc., Chem. Commun.* **1987**, 153; (b) Moody, C. M.; Young, D. W. *Tetrahedron Lett.* **1993**, *34*, 4667; (c) Exquerra, J.; Pedregal, C.; Micó, I.; Nájera, C. *Tetrahedron: Asymmetry* **1994**, *5*, 921; (d) Dieterich, P.; Young, D. W. *Org. Biomol. Chem.* **2006**, *4*, 1492.
- (a) For our recent review on the cinchona alkaloid-based phase-transfer catalysts for asymmetric synthesis, see: Jew, S.-s.; Park, H.-g. *Chem. Commun.* **2009**, 7090; For our recent reports on asymmetric phase-transfer catalysis, see: Yoo, M.-S.; Jeong, B.-S.; Lee, J.-H.; Park, H.-g.; Jew, S.-s. *Org. Lett.* **2005**, *7*, 1129; (b) Kim, T.-S.; Lee, Y.-J.; Jeong, B.-S.; Park, H.-g.; Jew, S.-s. *J. Org. Chem.* **2006**, *71*, 8276; (c) Lee, J.; Kim, M.-h.; Jew, S.-s.; Park, H.-g.; Jeong, B.-S. *Chem. Commun.* **2008**, 1932; (d) Kim, T.-S.; Lee, Y.-J.; Lee, K.; Jeong, B.-S.; Park, H.-g.; Jew, S.-s. *Synlett* **2009**, 671; (e) Kim, M.-h.; Choi, S.-H.; Lee, Y.-J.; Lee, J.; Nahm, K.; Jeong, B.-S.; Park, H.-g.; Jew, S.-s. *Chem. Commun.* **2009**, 782; (f) Park, Y.; Kang, S.; Lee, Y.-J.; Kim, T.-S.; Jeong, B.-S.; Park, H.-g.; Jew, S.-s. *Org. Lett.* **2009**, *11*, 3738.
- Ramachandran, P. V.; Madhi, S.; Bland-Berry, L.; Reddy, M. V. R.; O'Donnell, M. J. *J. Am. Chem. Soc.* **2005**, *127*, 13450.
- Byun, H.-S.; Reddy, K. C.; Bittman, R. *Tetrahedron Lett.* **1994**, *35*, 1371.
- (a) O'Donnell, M. J.; Bennett, W. D.; Wu, S. *J. Am. Chem. Soc.* **1989**, *111*, 2353; (b) Lygo, B.; Wainwright, P. G. *Tetrahedron Lett.* **1997**, *38*, 8595; (c) Corey, E. J.; Xu, F.; Noe, M. C. *J. Am. Chem. Soc.* **1997**, *119*, 12414; (d) Jew, S.-s.; Yoo, M.-S.; Jeong, B.-S.; Park, H.-g. *Org. Lett.* **2002**, *4*, 4245.